

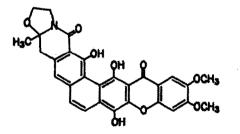
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Total Synthesis of Novel Xanthone Antibiotics (\pm)-Cervinomycins A_1 and A_2

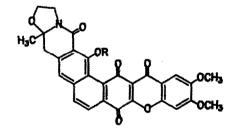
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Abstract: A total synthesis of novel heptacyclic antibiotics cervinomycin $A_1 \perp$ and $A_2 \geq$ following a convergent approach is reported. The cornerstone of our strategy was the construction of the central ring D through photochemical electrocyclization. The oxazolo-isoquinolinone fragment (ABC rings) $\underline{7}$ and the xanthone fragment (EFG rings) $\underline{8}$ were assembled through relatively straightforward synthetic protocols and coupled through a Wittig reaction to give $\underline{6}$ and set upthe key photocyclization. Our successful approach to $\underline{1}$ and $\underline{2}$ can be readily adapted to the synthesis of analogues of these interesting antibiotics.

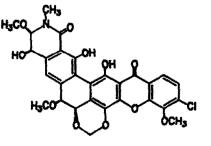
The genus streptomyces is a prodigious source of structurally variegated and biologically potent metabolites.¹ A new species <u>Streptomycis</u> <u>cervinus</u> sp.nov. in this genus was recently discovered and a collaborative effort led to the isolation and structure determination of novel antibiotic substances cervinomycins A₁ <u>1</u> and A₂ <u>2</u> in 1986.² These compounds



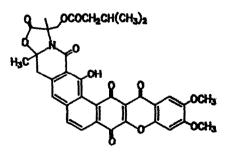
Cervinomycln A, 1



Cervinomycin A2 2



Lysolipin I 3



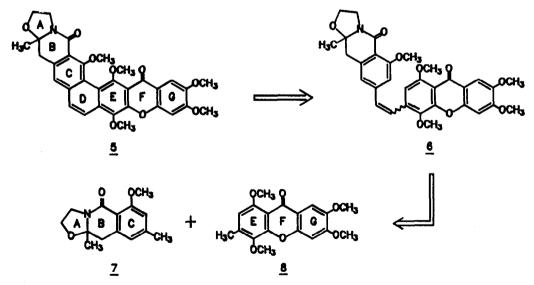
Citreamicin a 4

closely related as a redox pair, attracted immediate attention on account of their promising activity against anaerobic bacteria, mycoplasma and some gram positive bacteria and their structural novelty.

The cervinomycins A_1 and A_2 belong to a small but esoteric group of antibotics embodying an angularly fused polycyclic framework in which xanthone and isoquinolinone moieties are readily discernible.³ Some other members of this group that have been characterized are lysolipin $\underline{3}^{3b}$ and citreamicin $\underline{4}$.^{3e} The lure of novel structural features present in $\underline{1}-\underline{4}$ and their promising biological profile has generated considerable synthetic interest in these compounds. Initial model studies⁴ towards their synthesis have been followed by the first total synthesis of cervinomycin in 1989.⁵ Subsequently, two more total syntheses by us^{6a} and others,^{6b} following different strategies, have been reported. In addition, some more model studies have also been reported, indicating continued synthetic interest in these molecules.^{6C},^d We describe herein full details of our synthesis of 1 and 2 following a short convergent strategy.⁷

Synthetic Strategy

We conceived a synthetic approach to the heptacyclic framework of $\underline{1}$ and $\underline{2}$ through the protected derivative $\underline{5}$ which could be assembled through the union of two preformed ABC and EFG fragments $\underline{7}$ and $\underline{8}$, respectively. The retrosynthetic analysis (Scheme 1), indicated the synthetic plan that required parallel synthesis of the oxazolo-isoquinolinone $\underline{7}$ and the pentasubstituted xanthone $\underline{8}$ parts and their coupling to set up the formation of the strained ring D via photocyclization in $\underline{6}$. Interestingly, this



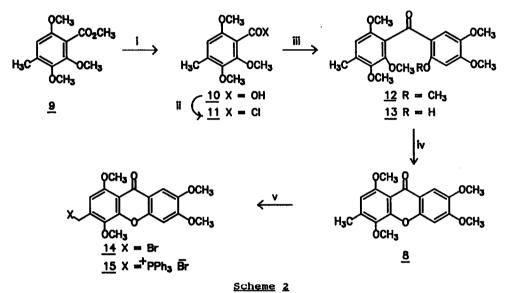
Scheme 1

strategy is not only economical and conceptually appealing but requires that the dense functionalization of $\underline{1}$ and $\underline{2}$ is built separately in fragments ABC and EFG and the cumbersome D ring, providing the angular fusion,

is generated only towards the end. For the coupling of ABC and EFG components to access the precursor $\underline{6}$ for photocyclization, Wittig reaction appeared most appropriate and complementary functionalization for this purpose was sought to be created through the benzylic methyl groups in $\underline{7}$ and $\underline{8}$.

Synthesis of the EFG Ring Fragment

While the number of synthetic and natural xanthones abound in the literature, none having the substitution pattern present in the EFG fragment $\underline{8}$ with a methyl group at the desired position has been reported. From the available options for the construction of $\underline{8}$, it was considered most convenient to access the xanthone framework <u>via</u> a base catalyzed cyclization of the corresponding benzophenone <u>13</u>. A synthesis of <u>13</u> from readily available starting materials and reagents is depicted in Scheme 2. The

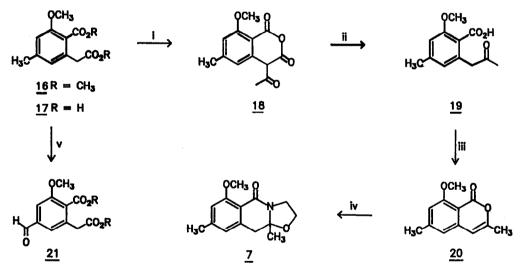


Reagents & Conditions: (i) 10% aq.KOH, CH₃OH, reflux, 4h, 95%; (ii) SOCl₂, reflux, 2h; (iii) 1,2,4-trimethoxybenzene, ether, anh.AlCl₃, r.t., 12h; (iv) CH₃OH, aq.K₂CO₃, reflux, 12h, 70%, (v) NBS, AIBN, CCl₄, reflux, 3h, 80%.

starting 2,3,6-trimethoxy-p-toluic acid methyl ester is available from orcinol in five straightforward steps,⁸ which were slightly modified to suit our requirements. The ester 9 was hydrolyzed to give the acid <u>10</u> which was further transformed to the acid chloride <u>11</u>. The acid chloride <u>11</u> was used to acylate 1,2,4-trimethoxybenzene in the presence of anhydrous AlCl₃ to furnish a mixture of hexamethoxybenzophenone <u>12</u> and its mono-demethylated derivative <u>13</u> in varying ratios depending upon the amount of the catalyst used and the time given.⁹ While <u>12</u> and <u>13</u> could be separated, in practice it was more convenient to subject the mixture to base catalyzed cyclization to yield xanthone <u>8</u> in good yield. The benzophenone <u>12</u> could be recovered unchanged from the reaction and could be recyclized <u>via</u> AlCl₃ mediated demethylation to <u>13</u>. The methyl group in xanthone <u>8</u> could be readily activated through benzylic bromination with N-bromosuccinimide (NBS) to furnish <u>14</u>. The bromomethyl group in <u>14</u> was then transformed to the required triphenylphosphonium bromide <u>15</u>, Scheme 2, the Wittig component for the contemplated coupling with the ABC fragment as per the retrosynthetic protocols shown in Scheme 1.

Synthesis of the ABC Ring Fragment

For the synthesis of the ABC portion of the cervinomycins, we chose to employ 3-methoxy-5-methylhomophthalic acid 17,¹⁰ readily obtainable in gram quantities from the Diels-Alder reaction between 6-methoxy4-methyl-2-pyrone and 1,3-dicarbomethoxyallene and hydrolysis of the resulting ester <u>16</u>. The acid <u>17</u> was converted¹¹ to isochroman-1,3-dione derivative <u>18</u> with Ac₂O-pyridine and further decarboxylated with aqueous base to give 2-acetylmethyl-6-methoxy-4-methylbenzoic acid <u>19</u>. Cyclization of <u>19</u> in the presence of Ac₂O and cat.HClO₄¹² gave 3,6-dimethyl-8-methoxyisocoumarin <u>20</u>. Treatment of <u>20</u> with 2-aminoethanol led to the oxazolo-isoquinolinone <u>7</u> with a quaternary methyl group at the ring junction in 50% yield, Scheme 3. Acquisition of the ABC fragment was revealed by its distinctive ¹H NMR spectrum (δ 1.27, 3H, s; 2.93, 1H, d and 3.08 1H, d).



Scheme 3

<u>Reagents & Conditions</u>: (i) Ac_2O , Py, 73% (ii) 10% aq.NaOH, ; (iii) Ac_2O , cat.HClO₄ 77%; (iv) 2-aminoethanol, CH₃OH, 50%; (v) NBS-CCl₄, [N(n-Bu)₄]₂Cr₂O₇-CHCl₃, 50%.

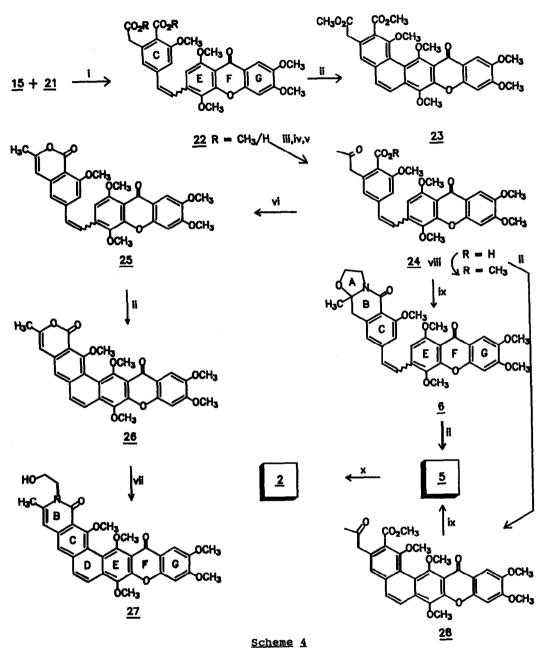
While the smooth attainment of $\underline{7}$ was gratifying, it was crucial at this stage to functionalize the benzylic methyl group present in $\underline{7}$ to enable its coupling with the ylide derived from <u>8</u>. However, the attempts to functionalize $\underline{7}$ with NBS, SeO₂, etc. at the methyl group proved troublesome as regioselectivity between the two benzylic positions in $\underline{7}$ could not be achieved. We, therefore, sought to revert to functionalizing the benzylic methyl group in 20. However, 20 also exhibited undesired regioselectivity and, on reaction with NBS, only allylic bromination was observed. These difficulties necessitated recourse to some tactical modifications and the successful strategy that emerged is as below.

Elaboration to Cervinomycins A_1 and A_2 2

Since it became difficult to prepare appropriately functionalized derivative of 7 for the ABC + EFG coupling, we sought to deploy a suitable C ring precursor endowed with a functionalized methyl group for the Wittig coupling with the EFG fragment 15. In this tactical manoeuvre, the construction of the rings A and B needed to be deferred till the end stage of the synthesis. Thus, the dimethylhomophthalate derivative 16 was converted to the bromomethyl derivative regioselectivley and was further oxidized to the aldehyde 21 using bis-tetrabutylammonium dichromate¹³ in good yield. The aldehyde 21 was then subjected to the Wittig reaction with the phosphonium ylide 15 derived from 14 in the presence of potassium carbonate as a base and 18-crown-6 as a PTC to furnish the coupled product 22, Scheme 4. The ¹H NMR spectral data for 22 showed it to be a mixture of E,Z-isomers but it was not considered necessary to separate them at this stage and the mixture was as such irradiated from a 450 W Hg lamp to furnish 23 in 20% yield. While the formation of CDEFG ring system 23 in a regioselective manner was a satisfying outcome and fortified our confidence in the overall synthetic plan, the yield was quite unacceptable for completing the rest of the synthesis which required elaboration of functionalities to install AB rings. Consequently, it was decided that the oxazolo-isoquinolinone moiety be constructed prior to the photocylization step.

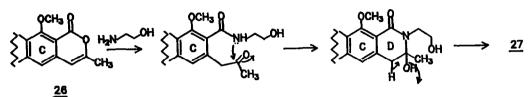
The Wittig product 22 was subjected to a three step sequence similar to that outlined in Scheme 3, to furnish the homologated keto acid 24 (R=H) which was characterized as the corresponding methyl ester. The acid 24 on treatment with acetic anhydride and $HClO_4$ (cat.) delivered the isocoumarin 25 in a good yield. The presence of vinylic methyl resonance at δ 2.25 was fully supportive of the formation of 25. The isocoumarin 25 on irradiation in the presence of iodine lead to the hexacyclic isocoumarin 26, in better yield than was observed for the photocyclization of the homophthalate 22. At this stage, the final formation of the AB rings was just a step away as demonstrated in the model studies (Scheme 3, 20 ---- 7). However, construction of the AB rings in 26 proved to be elusive and resulted only in the formation of the hexacyclic ring A secologue 27 of cervinomycin on reaction with 2-aminoethanol. Formation of 27 can be rationalized as shown in Scheme 5. Similar difficulties have been encountered previously.⁵ However, as to why <u>20</u> and <u>26</u> exhibit this difference in reactivity is not quite clear.

A search for the redressal of this unexpected hurdle at the threshold of our target made us to turn back and look at the precursor keto ester $\underline{24}$ (R=CH₃), which too can lead to the target compound on treatment with 2aminoethanol through a double cyclization sequence.⁶ Thus, the E,Z-mixture of the keto ester $\underline{24}$ was reacted with 2-aminoethanol and the oxazolo-isoquinolinone moiety was formed quite uneventfully as revealed by the appea-



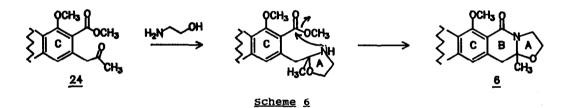
<u>Reagents & Conditions</u>: (i) K_2CO_3 , 18-crown-6, THF, 90%; (ii) hv, I₂, CH_2Cl_2 , 20-30%; (iii) 20% aq.KOH, CH_3OH , 90%; (iv) Ac_2o , DMAP, quant.; (v) 10% aq.NaOH, quant.; (vi) Ac_2O , $HClO_4$, $CHCl_3$, 75%; (vii) (a) 2-aminoethanol, CH_3OH ; (b) $BF_3.Et_2O$, CH_2Cl_2 , 70%; (viii) CH_2N_2 , Et_2O/CH_2Cl_2 , 85%; (ix) 2-aminoethanol, MeOH/ CH_2Cl_2 , 40-60%; (x) Ag_2O , 6N HNO₃, dioxan, 67%.

rance of the expected quarternary methyl singlet and benzylic methylene resonances in the ¹H NMR spectrum of <u>6</u>. Formation of <u>6</u> from <u>24</u> can be rationalized as shown in Scheme 6. The resulting ABC_DEFG precursor <u>6</u> on short exposure to pyrex filtered UV irradiation in the presence of a catalytic amount of iodine delivered cervinomycin A₁ trimethyl ether <u>5</u>. Alternatively, <u>5</u> could also be accessed from <u>24 via</u> first photocyclization to <u>28</u> followed by the treatment with 2-aminoethanol.



Scheme 5

Cervinomycin A_1 trimethyl ether 5 was easily converted to cervinomycin A_2 methyl ether 2 (R=CH₃) through oxidation with either silver oxide or ceric ammonium nitrate¹⁴ under carefully controlled conditions. The spectral data for cervinomycin A_2 methyl ether 2 was found to be identical with that reported in the literature.^{2C} The monomethyl ether 2 has already been converted to 2 (R=H) and $1.6^{6b,5}$ The present work, therefore, constitutes a total synthesis of cervinomycin A_1 and A_2 .



Experimental

Melting points were recorded on a Buchi SMP-20 apparatus and are uncorrected. IR spectra were recorded on a Perkin-Elmer 1310 spectrophotometer and were calibrated against the polystyrene absorption at 1601 cm⁻¹. Solid samples were recorded as KBr wafers. ¹H NMR (100 MHz) and ¹³C NMR (25 MHz) spectra were recorded on a JEOL FX 100 spectrometer unless and otherwise specified. NMR samples were dissolved in CHCl3-d solvent and chemical shifts are reported in δ scale relative to tetramethylsilane as the internal standard. Elemental analyses were performed on a Perkin-Elmer 240C CHN analyser. Reactions were monitored by TLC employing appropriate solvent systems. Moisture sensitive reactions were carried out by using the standard syringe-septum technique. Solvents for reactions were dried over appropriate drying agents. Usual work-up procedure includes washing of solvent extracts with water and brine, drying over anhydrous Na2SO4 and concentration at reduced pressure on a Buchi RE-121 rotary evaporator. Yields reported are isolated yields of homogeneous material judged by TLC and NMR spectroscopy. All irradiations were carried out using 450 W medium

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pressure Hanovia Hg immersion lamp.
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2,3,6-Trimethoxy-p-toluic acid methyl ester 9

2,3,6-Trihydroxy-p-toluic acid methyl ester⁸ (1.0g, 5 mmol) was dissolved in dry acetone (50 ml) to which anhydrous K_2CO_3 (5g, 36 mmol) and methyl iodide (7.0g, 50 mmol) were added and the resulting mixture was refluxed for 24h. The solvent was removed and the residue was diluted with water. Usual work-up with ether (3 x 50 ml) and concentration of the extracts gave the residue which was purified over SiO₂-gel column using 15% ethyl acetate/hexane mixture to yield 9 (1.06g, 88%); IR (neat): 1730, 1600, 1275, 1235, 1205 cm⁻¹; ¹H NMR: δ 6.44 (1H, s), 3.90 (3H, s), 3.88 (3H, s), 3.76 (6H, s), 2.26 (3H, s); Anal. Calcd. for $C_{12}H_{16}O_5$: C, 59.99; H, 6.71. Found: C, 59.34; H, 6.67.

3-Nethyl-1,4,6,7-tetramethoxyxanthone 8

To a solution of <u>9</u> (1.06q, 4.4 mmol) in CH₃OH (10 ml), 10% aqueous KOH solution (10 ml) was added and the mixture was refluxed with stirring for 4h. Methanol was removed under reduced pressure and contents were diluted with water and acidified (pH1) with dil.HCl. Extraction with ethyl acetate (3 x 20 ml) and removal of solvent gave 2,3,6-trimethoxy-4-methyl benzoic acid (0.95g, 95%); m.p. 154°C; IR (KBr): 2850(br), 1690, 1590, 1400, 1100, 920 cm^{-1} . The acid was refluxed with excess of thionyl chloride (10 ml) for 2h and excess thionyl chloride and HCl were completely removed under vacuum. The formation of 3,6-trimethoxy-4-methylbenzoyl chloride <u>11</u> was confirmed by IR (neat): 1780(s), 1600, 1120, 790 cm^{-1} . The acid chloride <u>11</u> was transferred using dry ether (15 ml) to a two necked RB flask containing 1,2,4-trimethoxybenzene (0.775g, 4.6 mmol) in dry ether (15 ml). To this mixture, anhydrous AlCl₃ (1.5g, 11 mmol) was added portion wise during 30 min. with the help of a solid addition funnel with stirring. The resulting solution was stirred at room temp. for 18h and poured onto crushed ice containing dil. HCl. Usual workup with ether and removal of solvent left a residue containing 2,3,6,2',4',5'-hexamethoxy-4-methylbenzophenone <u>12</u> and its monodemethylated derivative <u>13</u>. These were separated through SiO2-gel chromatography and characterized. <u>12</u>: IR (neat): 1730, 1620, 1400, 1030 cm⁻¹; ¹H NMR: δ 2.28 (3H, s), 3.59 (3H, s), 3.63 (3H, s), 3.72 (3H, s), 3.78 (3H, s), 3.85 (3H, s), 3.92 (3H, s), 6.42 (1H, s), 6.44 (1H, s), 7.42 (1H, s). <u>13</u>: IR (neat): 3300, 1730, 1600, 1220, 1040 cm⁻¹; ¹H NMR: δ 2.32 (3H, s), 3.47 (3H, s), 3.69 (3H, s), 3.85 (6H, s), 3.94 (3H, s), 6.08 (1H, s), 6.47 (1H, s), 6.92 (1H, s), 12.08 (1H, s).

The mixture of benzophenones <u>12</u> and <u>13</u> was dissolved in CH_3OH (20 ml) and potassium carbonate (5g) in water (45 ml) was added. The resulting mixture was refluxed overnight. Usual work-up with $CHCl_3$ (3 x 50 ml) gave a residue which was chromatographed on SiO_2 -gel using 80% ethyl acetate/hexane mixture to give the xanthone <u>8</u> (0.78g, 56%). The recovered benzophenone <u>12</u> was treated with anhydrous $AlCl_3$ in dry ether to demethylate a methoxy group on the ortho positions and repetition of the base catalyzed cyclization as described above gave an additional amount of <u>8</u> (0.2g), bringing the overall yield to 70%; m.p. 167-168°C; IR (KBr): 1650, 1620, 1600, 1490, 1270, 1120, 780 cm⁻¹; ¹H NMR: δ 2.42 (3H, s), 3.92 (3H, s), 3.97 (6H, s), 4.0 (3H, s), 6.56 (1H, s), 6.92 (1H, s), 7.62 (1H, s); ¹³C NMR: δ 16.35, 56.0, 56.12(2C), 60.83, 99.12, 105.40, 106.71, 111.12, 115.66, 137.36, 139.66, 146.6, 150.60, 151.00, 154.60, 155.48, 175.07. Anal. Calcd. for C₁₈H₁₈O₆: C, 65.44; H, 5.49. Found : C, 65.48; H, 5.50.

3-Bromomethyl-1,4,6,7-tetramethoxyxanthone 14

A mixture of 3-methyl-1,4,6,7-tetramethoxyxanthone <u>8</u> (0.25g, 0.75 mmol), CCl₄ (8 ml), NBS (0.15g, 0.87 mmol) and azo<u>bis</u>isobutyronitrile (AIBN, 0.02g) was refluxed under N₂ for 3h. The reaction mixture was concentrated and the residue chromatographed on SiO₂-gel using 70% ethyl acetate/hexane as an eluent to give <u>14</u> (0.248g, 80%); m.p. 206-208°C; IR (KBr): 1660, 1620, 1600, 1480, 1280, 780 cm⁻¹; ¹H NMR: δ 3.96 (3H, s), 3.98 (3H, s), 4.01 (3H, s), 4.08 (3H, s), 4.6 (2H, s), 6.72 (1H, s), 6.82 (1H, s), 7.6 (1H, s); ¹³C NMR: δ 26.65, 56.18, 56.42(2C), 61.77, 99.12, 105.54, 105.95, 113.12, 115.9, 136.13, 139.95, 147.0, 150.66, 155.07, 155.94, 174.95. Anal. Calcd. for C₁₈H₁₇BrO₆: C, 52.86; H, 4.19). Found: C, 52.90; H, 4.16.

1,4,6,7-Tetramethoxyxanthon-3-ylmethyltriphenylphosphonium bromide 15

A mixture of triphenylphosphine (0.24g, 0.9 mmol) and the bromomethyl xanthone <u>14</u> (0.25g, 0.6 mmol) in toluene (10 ml) was refluxed with stirring for 5h and allowed to cool to room temp. The precipitated phosphonium salt was filtered and washed with toluene (5 ml) and dried under vacuum to yield <u>15</u> (0.33g, 81%), m.p. 223-4°C.

3,6-Dimethyl-8-methoxyisocoumarin 20

To a well stirred mixture of acetic anhydride (1.5 ml) and dry pyridine (0.25 ml), 3-methoxy-5-methyl homophthalic $acid^{10}$ <u>17</u> (0.186g, 0.83 mmol) was added in small portions at room temp. and stirring was continued for 2h during which dry ether (5 ml) was added to facilitate smooth stirring. The solid product was filtered, washed with ether and dried under vacuum to give the intermediate 4-acetyl-8-methoxy-6-methylisochroman-1,3-dione <u>18</u> (0.15g, 73%) m.p. 182°C(d); IR (KBr): 1745, 1640, 1560, 1200, 1000, 840 cm⁻¹; ¹H NMR: δ 2.44 (3H, s), 2.56 (3H, s), 3.96 (3H, s), 6.68 (1H, s), 6.82 (1H, s).

To a stirred suspension of <u>18</u> (0.15g, 0.6 mmol) in water (1 ml) was added an aq. solution of sodium hydroxide (10%, 2 ml) and the resulting mixture was heated on a steam bath for 2h., cooled to room temp. and acidified with HCl to pH 1. Extraction with ethyl acetate (3x 7 ml) and addition of acetic anhydride (1 ml) and 70% perchloric acid (1 drop) resulted in a dark solution which was stirred under N₂ for 4h. The reaction was quenched with aq.NaHCO₃ and usual work-up gave a residue which was chromatographed using SiO₂-gel with 40% ethyl acetate/hexane as an eluent to give 3,6-dimethyl-8-methoxyisocoumarin <u>20</u> (0.95g, 77%) m.p. 137°C, IR (KBr): 1720, 1650, 1600, 1320, 1240, 1030, 960 cm⁻¹; ¹H NMR: δ 2.21 (3H, d, J=1Hz), 2.41 (3H, s), 3.96 (3H, s), 6.06 (1H, d, J=1Hz), 6.66 (2H, br s); ¹³C NMR: δ 19.12, 21.94, 55.89, 103.30, 106.18, 110.36, 117.19, 140.36, 146.95, 154.84, 159.65, 161.36. Anal. Calcd. for C₁₂H₁₂O₃: C, 70.57; H, 5.92. Found : C, 70.45; H, 5.90.

2,3,10,10a-Tetrahydro-8,10a-dimethyl-6-methoxyoxazolo[3.2-b]isoquinolin-5one 7

To a solution of 3,6-dimethyl-8-methoxyisocoumarin <u>20</u> (0.02g, 0.098 mmol) in CH₃OH (0.1 ml) was added 2-aminoethanol (0.1 ml) in CH₃OH and the mixture was stirred for 12h at room temp. Removal of solvent and chromatography on SiO₂-gel using ethyl acetate as an eluent afforded <u>7</u> (0.012g, 50%); m.p. 113°C, IR (KBr): 1655, 1610, 1310, 1100, 840 cm⁻¹; ¹H NMR: δ (500 MHz) 1.27 (3H, s), 2.35 (3H, s), 2.93 (1H, d, J=14.6Hz), 3.08 (1H, d, J=14.6Hz), 3.55 (1H, m), 3.90 (3H, s), 4.0-4.2 (3H, m), 6.62 (1H, s), 6.7 (1H, s); LRMS: m/z = 247 (M⁺, 30%), 162 (100%); Anal. Calcd. for C₁₄H₁₇NO₃: C, 67.99; H, 6.93; N, 5.66. Found: C, 68.06; H, 6.95. N, 5.70.

Dimethy1-5-formy1-3-methoxyhomophthalate 21

Dimethyl-3-methoxy-5-methylhomophthalate¹⁰ <u>16</u> (0.55g, 2.2 mmol), NBS (0.466g, 2.6 mmol) and AIBN (0.03g) in CCl₄ (10 ml) were refluxed under N₂ for 2h. Succinimide separated and was filtered and the residue after removal of solvent was chromatographed over SiO₂-gel column using 20% ethyl acetate/hexane as an eluent to give dimethyl-5-bromomethyl-3-methoxyhomophthalate (0.43g, 60%), m.p.88°C; IR (KBr): 2950, 1730, 1610, 1580, 1430, 1310, 1225, 1100 cm⁻¹; ¹H NMR: δ 3.64 (2H, s), 3.68 (3H, s), 3.85 (3H, s), 3.88 (3H, s), 4.44 (2H, s), 6.90 (2H, s); δ 32.24, 38.30, 51.59, 51.71, 55.59, 110.71, 123.07, 123.19, 133.42, 140.48, 156.95, 167.07, 170.48. Anal. Calcd. for C₁₃H₁₅BrO₅: C, 47.14; H, 4.57. Found: C, 47.41; H, 4.64.

The bromomethylhomophthalate (0.34g, 1.03 mmol) and <u>bis</u>-tetrabutylammoniumdichromate¹⁴ (1.5g, 2.07 mmol) were dissolved in $CHCl_3$ (7.0 ml) and the resulting solution was refluxed for 2h under N₂. The reaction mixture was filtered through florisil (60 mesh) and washed with ether. Combined organic washings were concentrated and the residue was chromatographed over SiO₂-gel employing 25% ethyl acetate/hexane to yield <u>21</u> (0.228g, 83%); m.p. 76°C; IR (KBr): 1720, 1695, 1590, 1460, 1280, 1220, 1100 cm⁻¹; ¹H NMR: δ 3.64 (2H, s), 3.65 (3H, s), 3.90 (3H, s), 3.92 (3H, s), 6.36 (1H, s), 6.37 (1H, s), 9.96 (1H, s); ¹³C NMR: δ 38.1, 51.83, 52.12, 55.89, 108.83, 125.54, 128.77, 133.9, 137.99, 157.30, 166.70, 170.30, 191.19; Anal. Calcd. for C₁₃H₁₄O₆: C, 58.64; H, 5.30 . Found: C, 58.72; H, 5.29.

(E, 2) - Dimethyl-5[(1,4,6,7-tetramethoxyxanthone-3-yl)ethen-2-yl]-3-methoxyhomophthalate 22

In a two necked RB flask were placed the Wittig salt <u>15</u> (0.225g, 0.33 mmol), anhydrous potassium carbonate (0.15g), 18-crown-6 (0.01g) and dry tetrahydrofuran (5 ml) and the suspension was stirred for 1h under N₂. To the resulting dark yellow solution, aldehyde <u>21</u> (0.098g, 0.369 mmol) in tetrahydrofuran (3 ml) was added dropwise. The reaction mixture was stirred at room temp. overnight and after the usual workup with CH_2Cl_2 furnished a residue which was purified by hydrolysis to the corresponding acid and re-esterification with diazomethane to give <u>22</u> (0.175g, 90%) as a mixture of E,Z-isomers; IR (KBr): 1735, 1620, 1600, 1470, 1425, 1290, 1120 cm⁻¹; ¹H NMR: δ 3.50-4.1 (m), 6.48(s), 6.7-7.64 (m); Anal. Calcd. for C31H30011: C, 64.35; H, 5.23. Found: C, 64.36; H, 5.23.

1,7,10,11,14-Pentamethoxy-3-(Methoxycarbonylmethyl)naphtho[1,2-b]xanthone-2-carboxylic acid methyl ester 23

The xanthone diester $\underline{22}$ (20 mg, 0.034 mmol) was irradiated in CH₂Cl₂ (40 ml) containing a catalytic amount of I₂ in a quartz vessel for 10 min. The solvent was removed and the residue was chromatographed on SiO₂-gel column using 1:1:1, CHCl₃:ethylacetate:hexane mixture as an eluent to furnish the pentacyclic diester $\underline{23}$ (4.0 mg, 20%); m.p. 206-209°C; IR (KBr): 1730, 1600, 1420, 1270, 1100 cm⁻¹; ¹H NMR: δ 3.48 (3H, s), 3.64 (3H, s), 3.72 (3H, s), 3.84 (2H, s), 3.96 (3H, s), 4.0 (3H, s), 4.04 (3H, s), 4.16 (3H, s), 7.00 (1H, s), 7.50 (1H, s), 7.66 (1H, d, J=9Hz), 7.72 (1H, s), 8.04 (1H, d, J=9Hz); [HRMS: M⁺ Calcd. for C₃₁H₂₈O₁₁: 576.1631. Found: 576.1661].

(E,2)-3-Methyl-8-methoxy-6-[(1,4,6,7-tetramethoxyxanthon-3-yl)ethen-2yl]isocoumarin 25

The diester 22 (0.163g, 0.28 mmol) in methanol (5 ml) was hydrolyzed to the corresponding homophthalic acid by refluxing with aq.KOH (10%, 5 ml) for 4h. Removal of methanol and acidification with HCl gave the homophthalic acid 22 (R=H) (0.115g, 0.28 mmol); IR (KBr): 3400, 1720, 1620, 1480, 1290 cm⁻¹. To a mixture of acetic anhydride (1 ml) and 4-dimethylaminopyridine (0.06g) was added homophthalic acid derivative 22 (0.155g, 0.28 mmol) and the mixture was stirred overnight at room temp. The yellow precipitates that separated were filtered and washed with ether and dried to give isochroman-1,3-dione derivative (0.16g, quantitative); IR (KBr): 1700, 1640, 1620, 1590, 1480 cm⁻¹. The above derivative was suspended in water (1 ml) and sodium hydroxide solution (10%, 2 ml) was added with stirring. The mixture was heated at 110°C for 2h and acidified to pH 1 with HCl. The precipitated product was filtered, washed with water (15 ml) and dried to give the keto acid 24 (R=H) (0.145g, 95%); IR (KBr): 1720, 1620, 1600, 1490, 1290, 1220, 1120 cm⁻¹.

To a stirred solution of the keto acid 24 (0.145g, 0.265 mmol) in CHCl₃ (30 ml) was added acetic anhydride (1.0 ml) followed by 70% perchloric acid (2 drops). Stirring was continued for 24h at room temp. and the solvent was removed. Addition of saturated NaHCO₃ (10 ml), extraction with CH₂Cl₂, usual work-up and removal of solvent gave a residue which was chromatographed on SiO₂-gel column using 2% CH₃OH/CHCl₃ to afford 25 (0.105g, 75%). Recrystallization furnished the major (E)-isomer, m.p. > 275; IR (KBr): 1740, 1650, 1620, 1600, 1470, 1220, 1200, 1120 cm⁻¹; ¹H NMR: δ 2.26 (3H, s), 3.98 (3H, s), 4.04 (6H, s), 4.07 (3H, s), 4.09 (3H, s), 6.18 (1H, s), 6.94 (2H, s), 7.04 (2H, br s), 7.22 (1H, d, J=16Hz), 7.62 (1H, d, J=16Hz), 7.64 (1H, s).

8,11,12,15,16-Pentamethoxy-3-methyl-isocoumarino[7,6-a]benzo[6,5-b]xanthone <u>26</u>

The diastereomeric mixture of isocoumarin-xanthone 25 (60 mg, 0.11 mmol) in CH₂Cl₂ (100 ml) containing catalytic amount of iodine was photolyzed in a quartz tube for 10 min. Solvent was removed and the residue was chromatographed over SiO₂-gel using 1:1:1

CHCl₃:ethylacetate:hexane solvent mixture as an eluent to give hexacyclic isocoumarin derivative <u>26</u> (20 mg, 33%); m.p. 190°C; IR (KBr): 1720, 1650, 1580, 1470, 1450, 1270, 1110, 1040 cm⁻¹; ¹H NMR: δ 2.26 (3H, d, J=1Hz), 3.40 (3H, s), 4.00 (3H, s), 4.06 (3H, s), 4.14 (3H, s), 4.20 (3H, s), 6.56 (1H, d, J=1Hz), 6.99 (1H, s), 7.16 (1H, s), 7.64 (1H, d, J=9Hz), 7.70 (1H, s), 8.18 (1H, d, J=9Hz); ¹³C NMR: δ (50 MHz) 19.90, 56.32, 56.51, 56.65, 61.88, 62.66, 99.40, 105.81, 106.21, 106.42, 108.8, 115.25, 115.71, 117.20, 119.55, 123.84, 129.52, 130.63, 137.64, 139.07, 139.78, 146.87, 147.69, 151.18, 152.86, 153.58, 155.48, 159.29, 160.68, 175.52; [HRMS: M⁺ Calcd. for C₃₀H₂₄O₉: 528.14202. Found: 528.1428].

Trimethoxycervinomycin A1 secologue 27

To a solution of hexacyclic isocoumarin $\underline{26}$ (10 mg, 0.019 mmol) in CH₃OH (1.0 ml) was added 2-aminoethanol (10 mg, 0.016 mmol) in CH₃OH and the mixture stirred at room temp. for 36h. Usual work-up with CH₂Cl₂ afforded a product which could not be characterized. However, brief exposure of this product to borontrifluoride-etherate in CH₂Cl₂ at 0°C afforded $\underline{27}$ (7 mg, 70%) m.p. 300°C; IR (KBr): 3350, 1650, 1620, 1600, 1580, 1480, 1430, 1300, 1280, 1200, 1050, 730 cm⁻¹; ¹H NMR: δ 2.42 (3H, s), 3.32 (3H, s), 4.0 (3H, s), 4.06 (3H, s), 4.14 (3H, s), 4.20 (3H, s), 4.0-4.5 (4H, m), 6.72 (1H, s), 7.0 (1H, s), 7.10 (1H, s), 7.66 (1H, d, J=9Hz), 7.7 (1H, s), 8.16 (1H, d, J=9Hz) [LRMS: m/z = 571 (M⁺, 100%)].

3-Acetylmethyl-1,7,10,11,14-pentamethoxynaphtho[1,2-b]xanthone-2-carboxylic acid methyl ester 28

The ketoacid 24 (R=H) (0.1g, 0.18 mmol) was taken in CH_2Cl_2 (5 ml) and ethereal diazomethane was added for esterification. The solvent was removed and the residue was chromatographed over SiO2-gel column using 2% CH₃OH/CHCl₃ as an eluent to furnish a mixture of (E,Z)-ketoester <u>24</u> (86 mg, 85%) which was irradiated in CH₂Cl₂ (200 ml) in the presence of catalytic I2 for 10 min. Solvent was removed and the residue was chromatographed using3:2:1 CHCl₂:ethyl acetate:hexanemixture as an eluent to give pentacyclic ketoester <u>28</u> (21 mg, 25%); m.p. 186°C; IR (KBr): 1720, 1650, 1610, 1580, 1480, 1460, 1430, 1280, 1110 cm⁻¹; ¹H NMR: δ 2.24 (3H, s), 3.50 (3H, s), 3.65 (3H, s), 3.90 (2H, s), 3.98 (3H, s), 4.01 (3H, s), 4.03 (3H, s), 4.18 (3H, s), 7.0 (1H, s), 7.4 (1H, s), 7.62 (1H, d, J=9.0Hz), 7.66 (1H, s), 8.02 (1H, d, J=9.0Hz); ¹³C NMR: δ 29.47, 47.94, 52.36, 56.24, 56.42, 60.77, 62.12, 62.30, 99.30, 106.01, 114.54, 115.83, 117.83, 119.71, 121.30, 124.48, 127.54, 130.18, 130.01, 131.54, 135.01, 137.83, 146.77, 147.71, 151.01, 155.31, 156.12, 157.07, 168.83, 175.36, 205.25; [LRMS: $m/z = 560 (M^+, 20\%), 529 (100\%)$].

(E,Z)-2,3,10,10a-Tetrahydro-6-methoxy-10a-methyl-8-[(1,4,6,7-tetramethoxyxanthon-3-yl)ethen-2-yl]oxazolo[3,2-b]isoquinolin-5-one 6

To a stirred solution of ketoester $\underline{24}$ (10 mg, 0.018 mmol) in CH₂Cl₂ (0.5 ml) was added 2-aminoethanol (10 mg, 0.16 mmol) in CH₃OH (0.2 ml) and the resulting mixture was stirred for 80h at room temp. The solvent was removed and the residue was chromatographed over SiO₂-gel using 2% CH₃OH/CH₃Cl to yield oxazolo-isoquinolinone derivative <u>6</u> (6.5 mg, 63%) as an (E,Z)-mixture; IR (KBr): 1640, 1620, 1600, 1480, 1430, 1400, 1290, 1220, 1120 cm⁻¹; [LRMS: m/z = 573 (M⁺, 80%), 558 (100%)].

Cervinomycin λ_1 -trimethylether 5

(a) By photolysis of <u>6</u>: The above product <u>6</u> was irradiated in CH_2Cl_2 (10 ml) containing catalytic of amount of I_2 in a quartz tube using pyrex filter, for 3 min. Solvent was removed and the residue was chromatographed over SiO₂-gel using 2% $CH_3OH/CHCl_3$ to give <u>5</u> (2.2 mg, 30%); m.p. > 300°C; IR (KBr): 1675, 1640, 1620, 1605, 1580, 1510, 1470, 1450, 1420, 1400, 1275, 1200, 1030, 1000 cm⁻¹; ¹H NMR: δ (400 MHz) 1.4 (3H, s), 3.21 (1H, d, J=14.9Hz), 3.29 (1H, d, J=14.9Hz), 3.65 (1H, m), 3.68 (3H, s), 3.70 (3H, s), 4.01 (3H, s), 4.06 (3H, s), 4.18 (3H, s), 4.16-4.24 (3H, m), 7.02 (1H, s), 7.35 (1H, s), 7.63 (1H, d, J=9Hz), 7.75 (1H, s), 8.08 (1H, d, J=9Hz); [LRMS: m/z = 571 (M⁺, 25%), 540 (100%)].

(b) From the pentacyclic ketoester <u>28</u>: To a solution of <u>28</u> (5 mg, 0.009 mmol) in CH₃OH (0.1 ml) was added 2-aminoethanol (7 mg, 0.11 mmol) in CH₃OH (0.1 ml). After stirring for 72 h at room temp., the solvent was removed and the residue was chromatographed over SiO_2 -gel to give <u>5</u> (2 mg, 40%) identical with that in the reaction described above.

Cervinomycin λ_2 -monomethyl ether 2

A mixture of cervinomycin A₁ trimethyl ether 5 (3 mg, 0.0053 mmol) and Ag₂O (0.5 mg) in dioxan (1 ml) was sonicated in the presence of a tiny drop of aqueous nitric acid (6N). The mixture was diluted with water and extracted with CH₂Cl₂ (3 x 5 ml). Usual work-up and chromatography over SiO₂-gel using 1% CH₃OH/CHCl₃ gave an orange crystalline compound identified as cervinomycin A₂ monomethyl ether 2 (R=CH₃, 2 mg, 67%); Alternately, 5 (4.5 mg, 0.0078 mmol) was dissolved in CH₂Cl₂ (0.5 ml) and CH₃CN (1.0 ml) was added with stirring followed by aq.ceric ammonium nitrate solution (10%, 0.5 ml). The resulting mixture was stirred for 5 minutes and the usual work-up followed by chromatography over SiO₂-gel gave 2 (R=CH₃, 4.3 mg, 93%); m.p. > 300°C; IR (KBr): 1690, 1650, 1615, 1450, 1270 cm⁻¹; ¹H NMR: δ 1.43 (3H, s), 3.27 (2H, d), 3.67 (1H, m), 4.01 (3H, s), 4.04 (3H, s), 4.21 (3H, s), 4.15-4.3 (3H, m), 7.16 (1H, s), 7.46 (1H, s), 7.66 (1H, s), 7.93 (1H, d, J=8.5Hz), 8.19 (1H, d, J=8.5Hz). The spectral data exactly matched those reported in the literature.

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References

- Glasby, J.S. <u>Encyclopaedia</u> of <u>Antibiotics</u>; John-Wiley & Sons (Chichester), 1979.
- (a) Omura, S.; Iwai, Y.; Hinotozawa, K.; Takahashi, Y.; Kato, J.; Nakagawa, A. <u>J. Antibiot.</u> 1982, <u>35</u>, 645. (b) Omura, S.; Nakagawa, A.; Kushida, K.; Lukacs, G. <u>J. Am. Chem. Soc.</u> 1986, <u>108</u>, 6088. (c) Nakagawa, A.; Omura, S.; Kushida, K.; Shimizu, H.; Lukacs, G. <u>J.</u>

Antibiot. 1987, 40, 301.

- (a) Albofungins: Gurevich, A.I.; Karapetyan, M.G.; Kolosov, M.N.; Omelchenko, V.N.; Unoprienko, V.I.; Petrenko, G.I.; Popravko, S.A. <u>Tetrahedron Lett.</u> 1972, 1751. (b) Lysolipins: Dobler, M.; Keller-Schierlein, W. <u>Helv. Chem. Acta</u> 1977, <u>60</u>, 178. (c) Simaomicins: Lee, T.M.; Carter, G.T.; Borders, D.B. J. <u>Chem. Soc. Chem. Commun.</u> 1989, 1771. (d) Actinoplanones: Kobayashi, K.; Nishino, C.; Ohya, J.; Sato, S.; Mikawa, T.; Shiobara, Y.; Kodama, M. J. <u>Antibiot.</u> 1988, <u>41</u>, 502, 741. (e) Citreamicins: Carter, G.T.; Niepsche, J.A.; Williams, B.R.; Borders, D.B. J. <u>Antibiotics</u> 1990, <u>43</u>, 504.
- 4. (a) Duthaler, R.O.; Heuberger, C.; Wegmann, U.H-U. <u>Chimia</u> 1985, <u>39</u>, 174. (b) Mehta,G.; Venkateswarlu, Y. <u>J. Chem. Soc. Chem. Commun.</u>
 1988, 1200. (c) Rao, A.V.R.; Reddy, K.K.; Yadav, J.S.; Singh, A.K. <u>Tetrahedron Lett.</u> 1988, <u>29</u>, 3991.
- Kelly, T.R.; Jagoe, C.T.; Li, Q. J. Am. Chem. Soc. 1989, 111, 4522.
 (a) Mehta, G.; Shah, S.R. Tetrahedron Lett. 1991, 38, 5195. (b) Rao, A.V.R.; Yadav, J.S.; Reddy, K.K.; Upender, V. Tetrahedron Lett. 1991, 38, 5199. (c) Deshpande, V.H.; Khan, R.A.; Rai, B.; Ayyangar, N.R. Abstracts, 17th IUPAC International Symposium on the Chemistry of Natural Products, New Delhi, 1990. (d) Jung, M.E.; Hagiwara, A. Tetrahedron Lett. 1991, 32, 3025.
- 7. For a preliminary communication of our work see ref.6a.
- Barton, D.H.R.; Bould, L.; Clive, D.L.J.; Magnus, P.D.; Hase, T. J. Chem. Soc. (C) 1971, 2204.
- 9. For an extensive study on acylation, demethylation and cyclization of benzophenones to xanthone see: Quillinan, A.J.; Scheinmann, F. J. <u>Chem. Soc. Perkin Trans. 1</u> 1973, 1329.
- Tamura, Y.; Fukata, F.; Tsugoshi, T.; Sasho, M.; Nakajima, Y.; Kita, Y.; <u>Chem. Pharm. Bull.</u> 1984, <u>32</u>, 3259.
- 11. Modi, A.R.; Usgaonkar, R.N. Indian J. Chem. 1979, 17B, 360.
- 12. Hauser F.M.; Rhee, R.P. J. Org. Chem. 1977, 42, 4155.
- 13. Landini, D.; Rolla, F. <u>Chem. Ind. (London)</u> 1979, 213.
- 14. (a) For similar oxidation with argentic oxide see: Snyder, C.B.; Rapoport, H.; <u>J. Am. Chem. Soc.</u> 1972, <u>94</u>, 227. (b) Jacob III, P.; Callery, P.S.; Shulgin, A.T.; Castagnoli Jr., N. <u>J. Org. Chem.</u> 1976, <u>41</u>, 3627.

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